

## **OPINION**

### **The presence of white blood cells in semen**

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## ABSTRACT

The present article critically reviews the recent data in the literature concerning the methods of quantitation of round cells in semen and in particular of white blood cells. The most reliable methods for the differentiation of round cells in semen are the peroxidase cytochemistry with the use of benzidine or o-toluidine and immunocytochemistry with the use of antibodies. The effect of increased numbers of white blood cells in semen on parameters of semen analysis as well as most of the sperm function tests is not yet clear. The clinical significance of such a finding is discussed along with the influence that it will have on fertility and the outcome of in vitro fertilization.

**Key words:** Peroxidase cytochemistry, semen, white blood cells

## INTRODUCTION

When assessing a semen analysis the number and type of round cells present should be taken into account along with "conventional" parameters, i.e. number, motility and morphology of spermatozoa (1). Round cells in semen can be distinguished into a) white blood cells (WBCs) and b) immature germ cells. Differentiation between these two cell types is considered of upmost importance for diagnostic and therapeutic purposes. For example, an increased number of WBCs in combination with other criteria may be indicative of a genital tract infection (20). On the other hand, an increased number of immature germ cells may be a sign of abnormal spermatogenesis (3).

Over the past few years immature germ cells have been the focus of attention due to the use of spermatids for in vitro conception. Round spermatid injection (ROSI) and round spermatid nuclear injection (ROSNI) were pioneered in Japan (4). These methods were the only therapeutic approach for some cases of azoospermia with spermatids taken from the testes (5) or the ejaculate (6). Identification of immature germ cells present in the ejaculate or the testicular tissue can be achieved with the use of antibodies (7) or it can be based on the morphological features of this cell category (8). Enriched fractions of spermatids can also be collected using cell separation techniques (8,9).

The injection of spermatids in the oocyte to achieve pregnancy has still obstacles to overcome (10), some of which are the correct identification as well as good quality of the cells isolated (11). The pregnancy rate after the use of round spermatids is disappointingly low (11) whereas the

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use of elongated spermatids gives acceptable pregnancy rates (12). This is not the only reason why the use of round spermatids warrants further attention. Very recent data have shown that the maturation of spermatozoa can only be arrested at the stage before meiosis (13). Large scale studies are needed to clarify the picture and decide on the future use of ROSI as a therapeutic approach.

### White blood cells in semen

There are two groups of WBCs in semen, those with granular cytoplasm and those with agranular cytoplasm. The former includes polymorphonuclear granulocytes, eosinophils and basophils, while the latter include monocytes and lymphocytes. Despite the presence of all above mentioned cells in semen samples, granulocytes - in particular polymorphonuclear granulocytes - are the predominant cell type (50-60%), macrophages follow (20-30%) and finally lymphocytes account for a very small proportion (2-5%). Other cell types are rare in semen samples (14-17). In a very interesting study (14) the number and type of cells present in the semen of large numbers of fertile and infertile men were counted using immunocytochemistry with antibodies specific for each cell type. This study revealed significant differences between the two groups of men. The number of WBCs in infertile men was significantly higher in comparison to the fertile group (median value: 1,035,000 per ejaculate versus 170,000 per ejaculate). The same was true concerning granulocytes (median value: 537,000 per ejaculate for the infertile group versus 100,000 per ejaculate for the fertile group). Significant increases were also observed in infertile men concerning the number of macrophages, B-lymphocytes and T-lymphocytes (14).

An issue widely discussed in the literature concerns the highest number of WBCs that can be considered "normal" for fertile semen. The WHO laboratory manual (18) considers 1 million/ml as the highest normal WBC's number. Most specialists have accepted this limit as a norm (3,14). In cases of accessory genital gland infection, the number of WBCs is usually higher

than the limit set in most studies, although there are a few papers that find no increase in the number of WBCs in infertile men with infection (16,17). An increased WBC number may be indicative of infection and it can affect the penetration of spermatozoa into the oocyte but it cannot be considered as the only indicator of the fertilizing potential of a semen sample (19).

### Methods for the determination of white blood cells in semen

It is generally recognized nowadays that round cells in semen samples must be counted accurately and distinguished carefully into WBCs and immature germ cells (20). Many methods have been developed to this purpose and the most commonly used ones are discussed below.

Bryan Leishman staining: This staining method was developed in 1976 by Couture (21). It uses anaphthol to differentiate between WBCs and immature germ cells. The method is simple, but may lead to an overestimation of the number of lymphocytes and underestimation of the number of granular cells, so it is not routinely used.

Elastase determination Elastase is an enzyme secreted by stimulated granulocytes. Its concentration in the seminal plasma can be measured with the use of Enzyme-Linked Immunosorbent Assay (ELISA) (22). This method correlates well with the immunohistochemical method (23) but not with the histochemical method of peroxidase stain (24). This discrepancy between the two methods can be explained by the fact that peroxidase staining measures a substance within the cytoplasm, whereas the enzyme elastase is secreted extracellularly. Elastase determination is a costly method demanding specialized equipment and for this reason it is not routinely used when studying WBCs in semen.

Papanicolaou staining: This is a widely used method for the evaluation of sperm morphology. The differentiation in this method is based on the shape and size of cell nuclei. However, cell classification is not always accurate with this method as polymorphonuclear granulocytes can be



taken for spermatids with lobular nuclei, and lymphocytes for monocytes (25).

Immunocytochemical method: Many monoclonal antibodies developed since 1980 can today be used to accurately identify all cell types, as there are antigens specific for each cell type (15, 26-28). This is the most accurate method available to date, but its high cost limits its use to research purposes, as it is too expensive to be used routinely.

Peroxidase cytochemistry: The original method developed by Endzt (29) was based on the benzidine compound for the differentiation between cell types. Later, Nahoum et al substituted o-toluidine for benzidine (30). The distinction between WBCs and immature germ cells is made possible because both benzidine and o-toluidine turn brown intracellularly due to the effect of peroxidase on hydrogen peroxide. As the method is both accurate and easy for an experienced scientist to perform and its cost relatively low, it is widely used routinely (20).

#### **White blood cells in semen and sperm parameters**

Many attempts have been made to answer the question of whether and to what extent WBCs - in particular polymorphonuclear granulocytes - can affect the quality of a semen sample. A number of studies have been conducted leading to varying results in an attempt to reach a conclusive answer (2,15,16,31,32). Older papers showed a decrease in number and motility of spermatozoa in infertile patients with leukocytospermia (15,31). Concerning the morphology of spermatozoa a reduction of normal forms was observed along with the decrease of their number and motility (32,33). These older findings have not been confirmed by recent studies (16,17) which involve large numbers of infertile men with leukocytospermia. On the contrary, most reports seem to agree that leukocytospermia has a negative effect on sperm function tests (34-39). The most commonly used tests are the following:

##### 1. Zona-free hamster egg penetration test (HEPT).

This test assesses the ability of spermatozoa for capacitation, acrosome reaction and fusion with the oolemma, which may not be present in acrosome-reacted sperm. In leukocytospermic samples this ability of spermatozoa was found reduced (34,35). These findings were experimentally confirmed by incubating semen samples with WBCs isolated from peripheral blood (36,37).

##### 2. Nuclear chromatin decondensation test (NCD).

This test assesses the capacity of nuclear chromatin to decondense after the influence of certain chemicals. A recent study showed a reduced capacity of nuclear chromatin to decondense in leukocytospermic semen samples (36).

##### 3. Hypo-osmotic swelling (HOS).

This test examines the ability of the sperm membrane for osmoregulation. Leukocytospermia has a negative effect on this test (38), but there is no general agreement on the issue (39).

In conclusion, it seems that leukocytospermia has a negative effect on sperm function tests that are related to the function of nuclear chromatin, the plasma membrane capacitation and acrosome reaction of spermatozoa.

#### **White blood cells in semen and fertility**

A number of studies have shown that the semen of infertile men contains higher numbers of WBCs (14,40,41). There are, however, papers with contradictory findings, i.e. a decrease in the number of WBCs in the semen of infertile men as compared to semen of fertile men (26,42). As already mentioned WHO suggests 1 million/ml as the highest number of WBCs in a normal semen sample. This generally accepted number is quite arbitrary and has, consequently, been questioned by several researchers (23,32,43,44). A particularly interesting study was conducted by Harrison et al (45) concerning this issue. They evaluated the number of WBCs in the semen of men who had fathered a child in the last twelve months. The numbers of WBCs found in these samples ranged from 0.5 - 16.5 million/ml. This



indicates that quite a high percentage of men who were definitely leukocytospermic, according to the WHO limit, were in fact fertile. This discrepancy puzzles researchers and there is a clear need to redefine relevant norms. It is also widely accepted that there is a need for an objective count of round cells in semen. WBCs should also be differentiated from immature germ cells using either the immunocytochemistry or the peroxidase cytochemistry method. This would ensure a clear differentiation between granular and agranular cells (peroxidase cytochemistry) or lead to the accurate identification of all cell types (immunocytochemistry).

### **White blood cells in semen and IVF outcome**

Does the number of WBCs in semen - in particular that of polymorphonuclear granulocytes - affect the outcome of IVF? A number of reports seem to suggest that there is a negative effect of WBCs on the outcome of IVF (46). Another study also confirms this negative effect and concludes that the effect is particularly detrimental if the number of WBCs exceeds 6 million/ml (47). As there is no change in conventional semen parameters, it can be concluded that an increased number of WBCs affects functional aspects of the spermatozoa, such as capacitation, hyperactivation and the acrosome reaction, thus preventing fertilization (48).

### **Reactive oxygen species and cytokines**

WBCs affect the fertilizing potential of the semen sample possibly through the action of the molecules that they secrete mainly reactive oxygen species (ROS) and cytokines. ROS are molecules such as the superoxide ion and the hydroxyl radical, which cause cell dysfunction. Cytokines are also a diverse group of molecules with various functions on cellular mechanisms. The presence of both groups was documented in semen over the past few years with yet undefined roles or modes of action.

ROS in the ejaculate are produced by WBCs (49-51) as well as by morphologically atypical spermatozoa (52). They can be measured in the seminal plasma with the use of chemiluminescence (49) or spectrophotometry (53). Increased amounts of ROS in semen have already been correlated with infertility (54,55). A number of sperm parameters as well as sperm function testing are affected by the presence of ROS (56-58).

ROS initiate the peroxidation of the unsaturated fatty acids of the sperm membrane, which alters fluidity affecting the membrane fusion necessary for fertilization (59). As a result increased ROS in seminal plasma can affect sperm function tests in vitro (60). They can also influence the outcome of in vitro fertilization and could be used as a predictive index (48).

The presence of cytokines in semen was first reported by Hill and his coworkers (61) and since then many research groups have attempted to clarify the role of cytokines in fertility. It has been reported that the levels of interleukin-2 in seminal plasma of infertile patients are decreased (62) whereas interleukin-11 levels are increased especially in the presence of urogenital infections (63). Patients with varicocele or infection have been found to have increased interleukin-6 levels in seminal plasma (64). Cytokines can also affect the motility of spermatozoa (65, 66) although this finding has not been confirmed by others (67, 68).

It becomes evident that consensus regarding the role of both ROS and cytokines in the ejaculate and the effect that they might have on the fertilizing potential of a semen sample has not yet been reached. There are many parameters that one has to consider such as the antioxidant systems of seminal plasma and synergy between different cytokines. Further research in the future will give definite answers.

### **What is the actual role of white blood cells in semen?**

Lately, systematic research has been initiated into the actual role of WBCs in semen (69,70). Aitken and Baker (71) in a recent article



humorously wonder whether WBCs in semen are passengers, terrorists or good Samaritans. In other words, the conventional view that the presence of WBCs in semen is definitely detrimental is being questioned. Contradictory data are accumulating with time (17,44). The conclusion so far is that assessment should take into account not only the population of polymorphonuclear granulocytes but also the immature germ cells, which may be responsible for negative effect, observed on certain sperm parameters (71).

In conclusion, although leukocytospermia is indicative of infection, it does not necessarily have a negative effect on the fertilizing capacity of a semen sample. However, it must be emphasized that an accurate count as well as differentiation of round cells in semen is strongly recommended, as this will be a valuable tool for both diagnostic and therapeutic decisions. That is particularly true when applying the latest methods of ROSNI and ROSI using round spermatids isolated from semen and then injected into oocytes (4,6), where counting and differentiation of round cells becomes imperative.

## REFERENCES

1. Yanushpolsky EH, Potitch JA, Hill JA and Anderson D J. Is leukocytospermia clinically relevant? *Fertil Steril* 1996; 66(5):822-825.
2. Comhaire F, Verscraegen G, Vermeulen L. Diagnosis of accessory gland infection and its possible role in male infertility. *Int J Androl* 1980;3:32-45.
3. Nieschlag E, Behre HM, Meschede D. Diseases of the seminal ducts. In *Andrology. Male Reproductive Health and Dysfunction*. Eds Nieschlag E, Behre H M. Springer, 1997.
4. Hannay T. New Japanese IVF method finally made available in Japan. *Nature Med* 1995; 1: 289-290
5. Gil-Salom M, Minguez Y, Rubio C, De los Santos M, Remohi J, Pellicier A. Efficacy of intracytoplasmic sperm injection using testicular spermatozoa. *Hum Reprod* 1995; 10(12): 3166-3170
6. Tesarik J, Mendosa C, Testart J. Viable embryos from injection of round spermatids into oocytes. *N. Engl J Med* 1995; 333: 525
7. Ezech UIO, Martin M, Cooke ID, Moore HDM. Correlation of testicular pathology and sperm extraction in azoospermic men with ejaculated spermatids detected by immunofluorescent localization. *Hum Reprod* 1998; 13(11): 3061-3065
8. Angelopoulos T, Krey L, McCullough A, Adler A, Grifo J. A simple and objective approach to identifying human round spermatids. *Hum Reprod* 1997; 12(10): 2208-2216
9. Aslam I, Robins A, Dowell K, Fishel S. Isolation, purification and assessment of viability of spermatogenic cells from testicular biopsies of azoospermic men. *Hum Reprod* 1998; 13(3):639-645
10. Sousa M, Barros A, Tesarik J. Current problems with spermatid conception *Hum Reprod* 1998; 13(2): 255-258
11. Vandezwalmen P, Zech H, Birkenfeld A. Intracytoplasmic injection of spermatids retrieved from testicular tissue: influence of testicular pathology, type of selected spermatids and oocyte activation. *Hum Reprod* 1997; 12: 1203-1213
12. Fishel S, Green S, Bishop M. Pregnancy after intracytoplasmic injection of spermatid. *Lancet* 1995;245:1641-1642
13. Silber SJ, Jonson L. Are spermatid injections of any clinical value? ROSNI and ROSI revisited. *Hum Reprod* 1998; 13(3): 509-523
14. Wolff H, Anderson DJ. Immunohistologic characterization and quantitation of leukocyte subpopulation in human semen. *Fertil Steril* 1988;49:497-504.
15. Eggert-Kruse W, Bellmann A, Rohr G, Tilgen W, Runnebaum B. Differentiation of round cells in semen by means of monoclonal antibodies and relationship with male fertility. *Fertil Steril* 1992;58:1046-1055.
16. Aitken RJ, West K., Buckingham D. Leukocytic infiltration in the human ejaculate and its association with semen quality, oxidative stress and sperm function. *J Androl* 1994;15:343-352.
17. Tomlinson MJ, Barratt CLR, Cooke ID. Prospective study of leukocytes and leukocyte subpopulations in semen suggests they are not a cause of male infertility. *Fertil Steril* 1993; 60:1069-1075.
18. World Health Organization (WHO) laboratory manual for the examination of human semen and sperm-cervical mucus inter-action. Cambridge: Cambridge University Press, 1992.
19. Van der Ven HH, Jeyendran RS, Perez-Pelaez M, Al-Hasani S, Diedrich K, Krebs D. Leucospermia and the fertilizing capacity of spermatozoa. *Eur J Obstet Gynecol Reprod Biol* 1987; 24: 49-52
20. Potitch JA, Wolff H, Hill JA, Anderson DJ. Comparison of methods to enumerate white blood cells in semen. *Fertil Steril* 1993;60(2):372-375.
21. Couture M, Ulstein M, Leonard J, Paulsen J R. Improved staining method for differentiating immature germ cells from white blood cells in human seminal fluid. *Andrologia* 1976; 8:61-66.
22. Jochum M, Papst W, Schill WB. Granulocyte elastase as a sensitive diagnostic parameter of silent male genital tract inflammation. *Andrologia* 1986;18:413-419.
23. Wolff H, Anderson DJ. Evaluation of granulocyte elastase as a seminal plasma marker for leukocytospermia. *Fertil Steril* 1989; 50:129-132.
24. Wolff H, Panhans A, Zebhauser M, Meurer M. Comparison of three methods to detect white blood cells in



- semen: Leukocyte esterase dipstick, granulocyte elastase enzyme immunoassay, and peroxidase cytochemistry *Fertil Steril* 1991;58(6):1260-1261.
25. Wolff H. The biologic significance of white blood cells in semen. *Fertil Steril* 1995;63(6):1143-1157.
  26. El-Demiry MIM, Young H, Elton RA, Hargreave TB, James K, Chisholm GD. Leucocytes in the ejaculate from fertile and infertile men. *Br J Urol* 1986;58:715-720.
  27. Harrison PE, Barratt CLR, Robinson AJ, Kessopoulou E, Cooke ID. Detection of white blood cell populations in the ejaculate of fertile men. *J Reprod Immunol* 1991;19:95-98.
  28. Schobel WA, Schieferstein G, Uchanska-Ziegler B. Immuno-cytochemical characterization of round cells in human semen using monoclonal antibodies and the APAAP-technique. *Andrologia* 1989;21:370-376.
  29. Endtz AW. Een methode om het vochtig urinesediment en het vochtige menlijke sperma rechtstreeks te kleuren. *Ned Tijdschr Geneesk* 1972;116:681-685.
  30. Nahoum CRD, Cardozo D. Staining for volumetric count of leukocytes in semen and prostate-vesicular fluid. *Fertil Steril* 1980;34(1):68-69.
  31. Talbert LM, Hammond MG, Halme J, O'Rand M, Fryer JG, Ekstrom RD. Semen parameters and fertilization of human oocytes in vitro: a multivariable analysis. *Fertil Steril* 1987;48:270-277.
  32. Gonzales GF, Kortebani G, Mazzoli AB. Leukocytospermia and function of the seminal vesicles on seminal quality. *Fertil Steril* 1992;57:1058-1065.
  33. Wolff H, Politch JA, Martinez A, Haimovici F, Hill JA, Anderson DJ. Leukocytospermia is associated with poor semen quality. *Fertil Steril* 1990;53:528-536.
  34. Berger RE, Karp LE, Williamson RA, Koehler J, Moore DA, Holmes KK. The relationship of pyospermia and seminal fluid bacteriology to sperm function as reflected in the sperm penetration assay. *Fertil Steril* 1982;37:557-564.
  35. Maruyama K, Hale RW, Rogers BJ. Effects of white blood cells on the in vitro penetration of zona-free hamster eggs by human spermatozoa. *J Androl* 1985;6:127-135.
  36. Vogelpoel FR, van Kooij RJ, te Velde ER, Verhoef J. Influence of polymorphonuclear granulocytes on the zona-free hamster oocyte assay. *Hum. Reprod.* 1991;6:1104-1107.
  37. Chacho KJ, Andersen PJ, Scommegna A. The effect of peritoneal macrophage incubates on the spermatozoa assay. *Fertil Steril* 1987;48:694-696.
  38. Gavella M, Lipovac V. Effect of leukocytes on the hypo-osmotic swelling test of human sperm. *Arch Androl* 1993;30:55-61.
  39. Chan PJ, Su BC, Tedway DR, Whitney EA, Pang SC, Corselli I, Jacobson JD. White blood cells in semen affect hyperactivation but not sperm membrane integrity in the head and tail regions. *Fertil Steril* 1994;61:986-989.
  40. Auroux M, Collin C, Couvillers ML. Do nonspermatozoal cells mainly stem from spermiogenesis? Study of 106 fertile and 102 subfertile men. *Arch Androl* 1985;14:73-80.
  41. Wang AW, Politch J, Anderson DJ. Leukocytospermia in male infertility patients in China. *Andrologia* 1994;26:167-172.
  42. Kung AWC, Ho PC, Wang C. Seminal leukocyte subpopulations and sperm function in fertile and infertile Chinese men. *Int J Androl* 1993;16:189-194.
  43. Tomlinson MJ, Barratt CLR, Bolton AE, Lenton EA, Roberts H B, Cooke ID. Round cells and sperm fertilizing capacity: the presence of immature germ cells but not seminal leukocytes are associated with reduced success of in vitro fertilization. *Fertil Steril* 1992;58:1257-1259.
  44. Tomlinson MJ, White A, Barratt CLR, Bolton AE, Cooke ID. The removal of morphologically abnormal sperm forms by phagocytes: a positive role for seminal leukocytes? *Hum Reprod* 1992;7/4:517-522.
  45. Harrison PA, Barrat CLR, Robinson AJ, Kessopoulou E, Cooke ID. Detection of white blood cell populations in the ejaculate of fertile men. *J Reprod Immunol* 1991;19:95-98.
  46. De Geyter C, De Geyter M, Behre HM, Schneider HPG, Nieschlag E. Peroxidase-positive cells and microorganisms in human semen together with antibiotic treatment adversely influence the out-come of in-vitro fertilization and embryo transfer. *Int J Androl* 1994;17:127-134.
  47. Cohen J, Edwards R, Fehilly C, Fishel S, Hewitt J, Purdy J. In vitro fertilization: a treatment for male infertility. *Fertil Steril* 1985;43:422-432.
  48. Sukcharoen N, Keith J, Irvine DS, Aitken J. Predicting the fertilizing potential of human suspensions in vitro: importance of sperm morphology and leukocyte contamination. *Fertil Steril* 1995;63(6):1293-1300.
  49. Krausz C, West K, Buchingham D, Aitken J. Development of a technique for monitoring the contamination of human semen samples with leukocytes. *Fertil Steril* 1992; 57/6 1317-1325.
  50. Plante M, Laminare E, Gagnon C. Reactive oxygen species released by activated neutrophils, but not by deficient spermatozoa, are sufficient to affect normal sperm motility. *Fertil Steril* 1994; 62: 387-393.
  51. Henkel R, Ichikawa T, Sanchez R, Miska W, Ohmori H, Schill W-B. Differentiation of ejaculates showing reactive oxygen species production by spermatozoa or leukocytes. *Andrologia* 1997;29: 295-301.
  52. Kessopoulou E, Tomlinson MJ, Barratt CLR, Bolton AE, Cooke ID. Origin of reactive oxygen species in human semen: spermatozoa or leukocytes? *J Reprod Fertil* 1992; 94: 463-470.
  53. Kovalski N., Lamirante E., Gagnon C. Determination of neutrophil concentration in semen by measurement of superoxide radical formation. *Fertil Steril* 1991; 56/5: 946-953.
  54. de Lamirande E, Leduc E, Iwasaki V, Hassouna M, Gagnon C. Increased reactive oxygen species formation in semen of patients with spinal cord injury. *Fertil Steril* 1995;63/3: 637-642.
  55. Mazzilli F, Rossi T, Marchesini M, Ronconi C, Dondero F. Superoxide anion in human semen related to seminal parameters and clinical aspects. *Fertil Steril* 1994; 62/4:862-868.
  56. Kurpisz M., Miesel R., Sanocka D., Jedrejczak P. Seminal plasma can be a predictive factor for male infertility. *Hum Reprod* 1996; 11/6: 1223-1226.

57. Aitken RJ, Buckingham DW, Brindle J, Gomez E, Baker GHW, Irvine SD. Analysis of sperm movement in relation to the oxidative stress created by leukocytes in washed sperm preparations and seminal plasma. *Hum Reprod* 1995; 10/8: 2061-2071
58. Fedder J. Nonsperm cells in human semen: with special reference to seminal leukocytes and their possible role in fertility. *Arch Androl* 1996; 36: 41-65
59. Aitken RJ, Clarkson JS. Cellular basis of defective sperm function and its association with the genesis of reactive oxygen species by human spermatozoa. *J Reprod Fertil* 1987; 81: 459-469
60. Aitken RJ, West KM. Analysis of the relationship between reactive oxygen species production and leukocyte infiltration in fractions of human semen separated on Percoll Gradients. *Int J Androl* 1990; 13:433-451
61. Hill JA, Haimovici F, Potich JA, Anderson DJ. Effects of soluble products of activated lymphocytes and macrophages (lymphokines and monokines) on human sperm motion parameters. *Fertil Steril* 1987; 47: 460-465
62. Naz RK, Evans L, Armstrong JS, Sikka SC. Decreased levels of interleukin-12 are not correlated with leukocyte concentration and superoxide dismutase activity in semen of infertile men. *Arch Androl* 1998;41:91-96
63. Metalliotakis I, Kyriakou D, Fragouli Y, Loutradis D, Goumenou A, Koumantakis E. Determination of interleukin-11 in seminal plasma and elevated IL-11 in seminal plasma of infertile patients with urogenital infection. *Arch Androl* 1998; 41:177-183
64. Zalata A, Hafez T, Hoecke MJV, Comhaire F. Evaluation of  $\beta$ -endorphin and interleukin-6 in seminal plasma of patients with certain andrological diseases. *Hum Reprod* 1995;10/12: 3161-3165
65. Eisermann J, Register KB, Strickler RC, Collins JL. The effect of tumor necrosis factor on human sperm motility in vitro. *J Androl* 1989;19:270-274
66. Estrada LS, Champion HC, Wang R, Rajasekaran M, Hellstrom WJG, Aggarwal B, Sikka SC. Effect of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interferon- $\gamma$  (IFN- $\gamma$ ) on human motility, viability and motion parameters. *Int J Androl* 1997;20: 237-242
67. Haney AF, Hughes SF, Weinberg JB. The lack of effect of tumor necrosis factor-alpha, interleukin-1-alpha and interferon-gamma on human sperm motility in vitro. *J Androl* 1992; 13: 249-253
68. Hussenet F, Dousset B, Cordonnier JL, Jacob C, Foliquet B, Grignon G, Nabet P. Tumor necrosis factor alpha and interleukin 2 in normal and infected seminal fluid. *Hum Reprod* 1993; 8: 409-411
69. Barratt CLR, Bolton AE, Cooke ID. Functional significance of white blood cells in the male and female reproductive tract. *Human Reprod* 1990;5(6):639-648.
70. Anderson DJ. Should male infertility patients be tested for Leukocytospermia? *Fertil Steril* 1995;63:246-248.
71. Aitken RJ, Baker GHW. Seminal leukocytes: passengers, terrorists or good Samaritans? *Hum Reprod* 1995;10/7:1736-1739.

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